# Circuit changes augment disinhibited shock responses in computer models of neocortex WW Lytton, SA Neymotin, HK Lee, DJ Uhlrich, AA Fenton 1,2,4

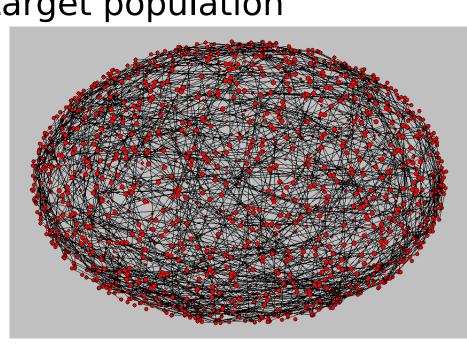
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There is limited knowledge and some disagreement regarding the density of connections between cortical layers. There is still less known about connectivity within each layer. In the absence of definite knowledge, modeling allows us to test hypotheses about cortical wiring and dynamics in-silico.

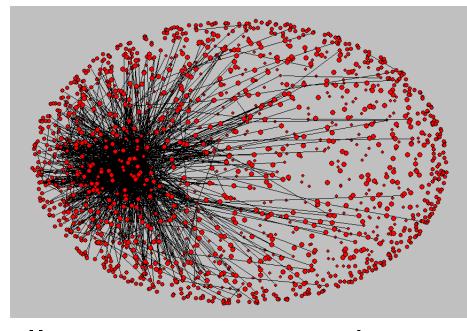
#### **Graph Methods**

Network setup: Graph measures will differ depending on how we connect cells within or between layers. Since the basic models give densities but not distributions, we tried 4 different distributions:

1. Uniform: each cell connects randomly to any other cell in target population



2. Scale-Free: most cells have few connections, but a few hubs connect to most other cells

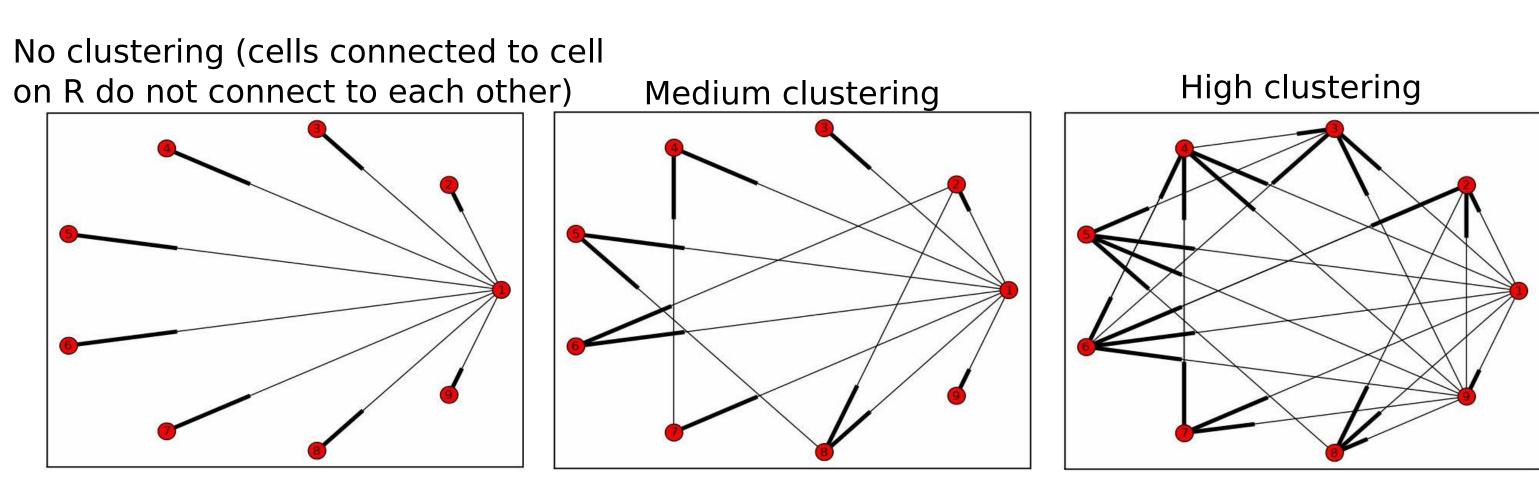


- 3. Fixed: Divergence is fixed for each cell; convergence varies.
- 4. Normal: Divergence follows normal distribution.

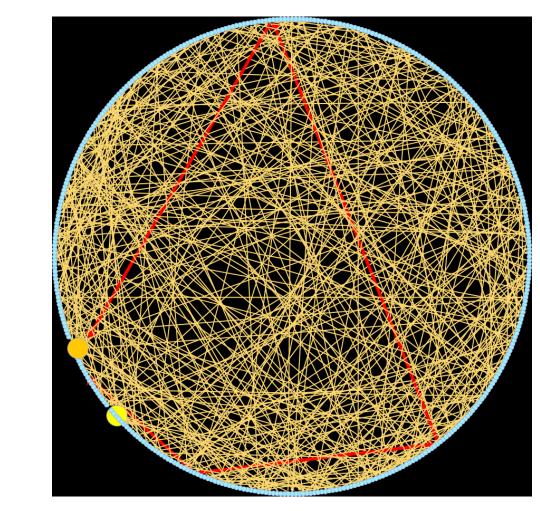
#### Measures:

Standard neurobiological connectivity measures -divergence, convergence, connection density -- are complemented by layer-specific or cell-specific graphtheoretic measures in order to characterize cortical wiring. 3 major graph-theoretic measures were used:

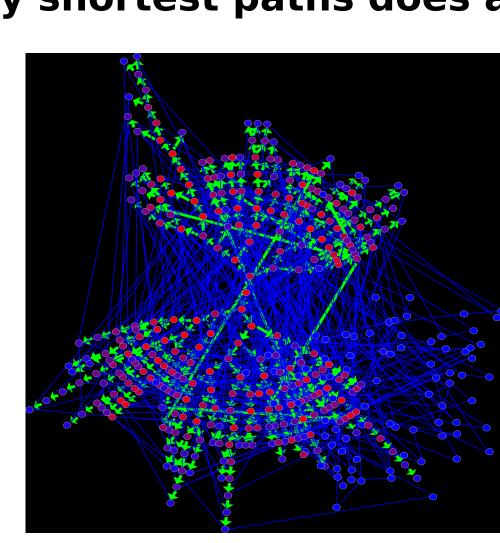
1. Clustering coefficient: How likely are neighbors of a cell to be connected to each other? A cluster is similar to a Hebb cell assembly.



2. Path length: average of shortest paths to connect two units. "Path" in graph theory is # of edges between nodes (cells) crossed to get from node A to node B. The path length shown below is 4.



3. Centrality: how many shortest paths does a cell lie on?

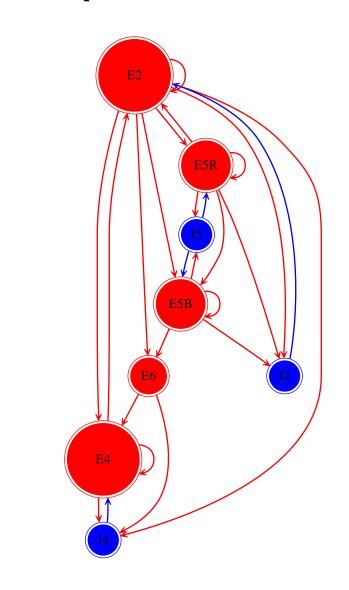


### 4 models all show Layer 2/3 dominance

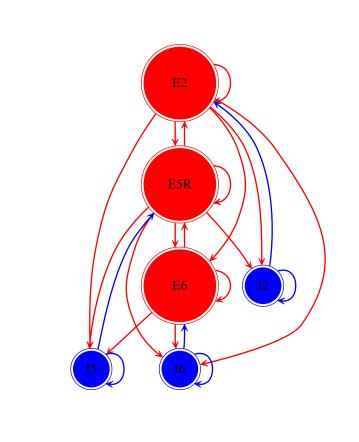
Automated graph analysis of 4 published cortical models. L2/3 (E2) is a central hub. Reciprocal excitatory feedback between L5 (E5) and L2/3 provide the next level of interaction for activating epileptiform activity.

Red=excitatory. Blue=inhibitory. Size of circles proportional to # of cells. Straightness of edges proportional to density of connections. Length of edges inversely proportional to density of connections. Circles sorted top to bottom by total average divergence.

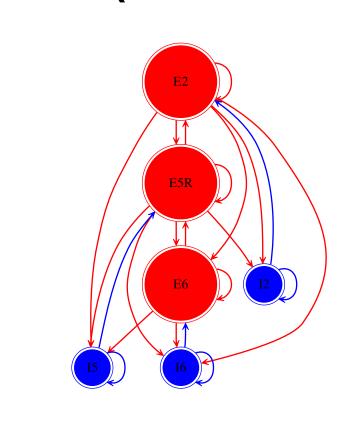
Primary Visual Cortex (based on Martin model [4,6,7,10])



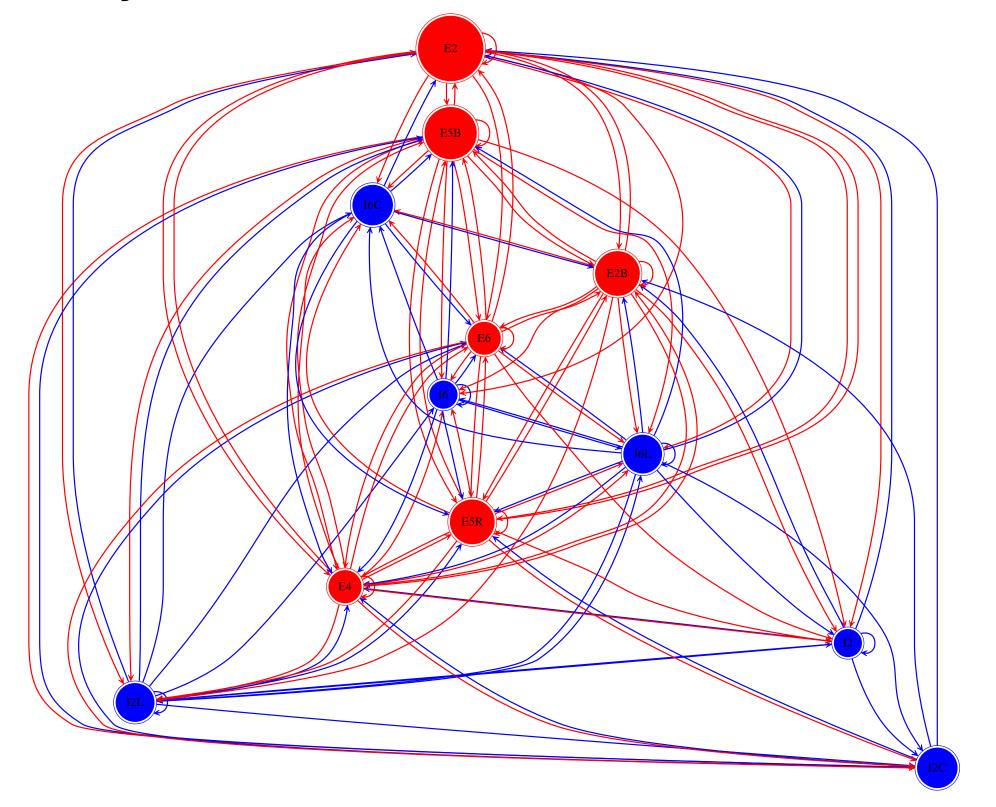
Primary Visual Cortex (based on Franaszczuk model [2])



Primary Motor Cortex (based on Tononi model [8,12])



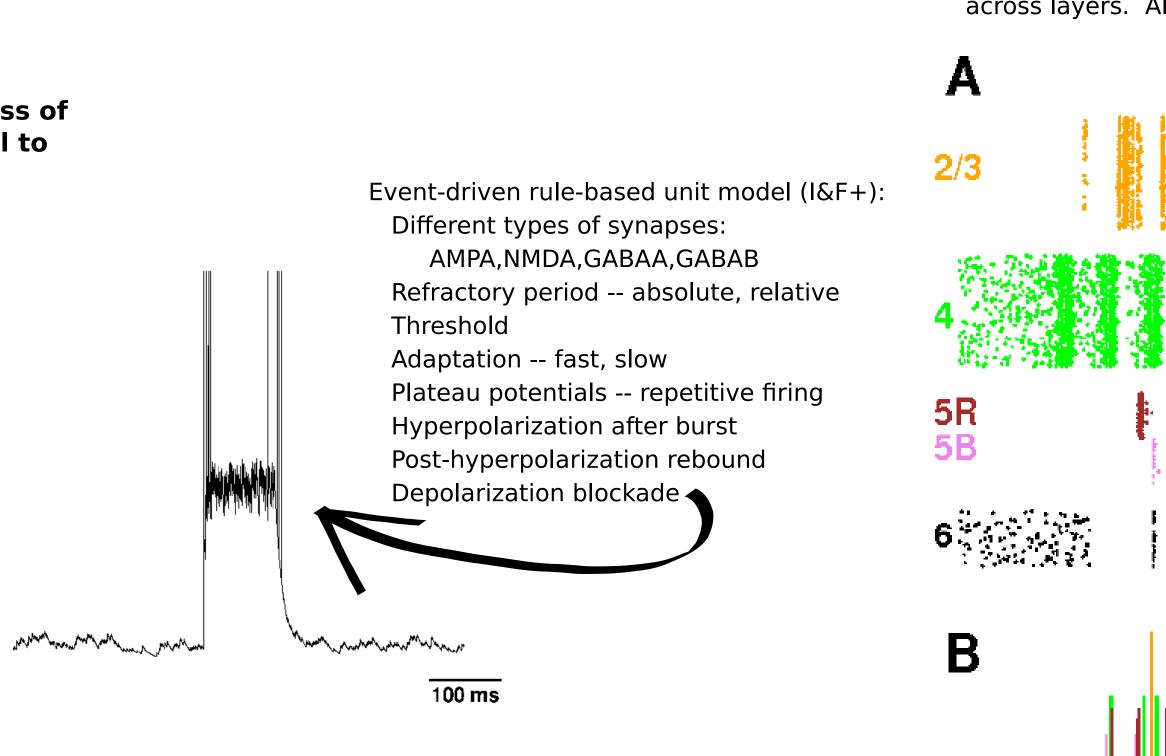
Auditory Cortical Column (based on Traub model [24])



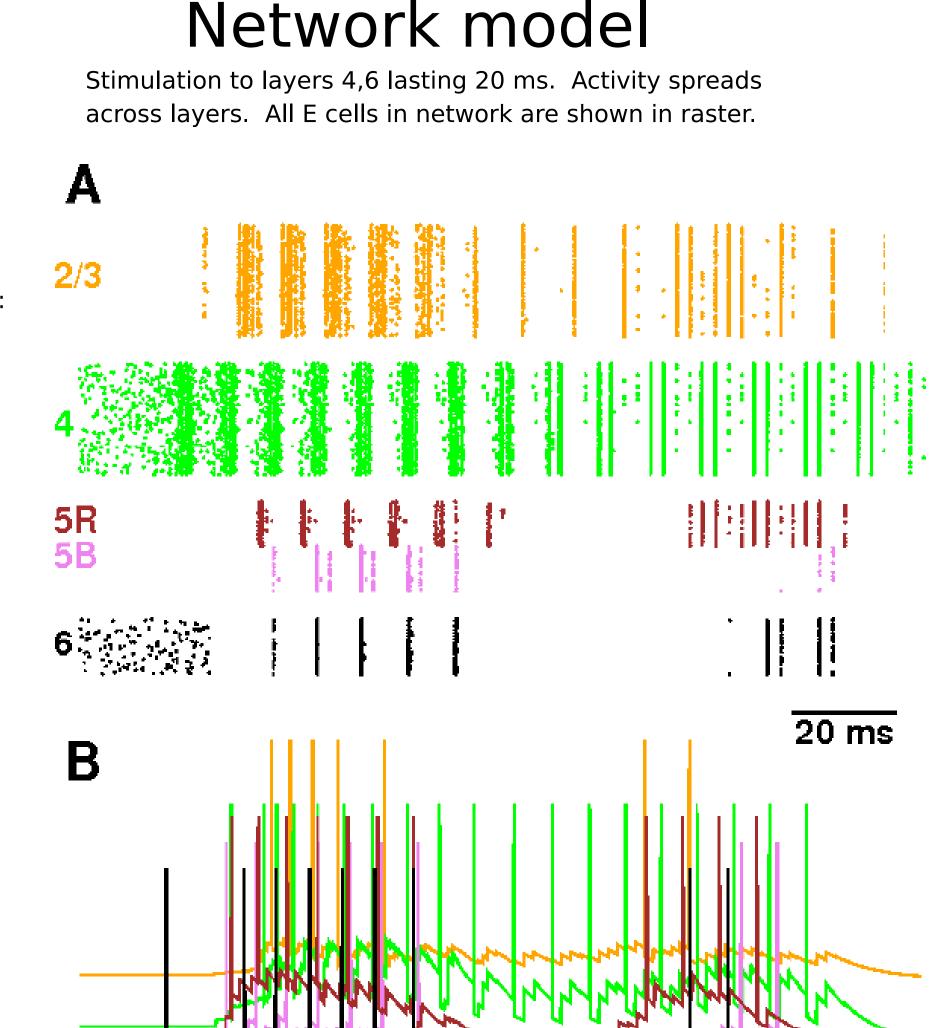
Acronym Key: E=excitatory; I=inhibitory; L=layer. E2 are excitatory cells in layer 2/3 (2 is short for L2/3); most E cells are pyramids; most I cells are basket cells; I2Q are double bouquet cells which may be in L2/3 or L5; I2C, I4, I6C are chandelier cells; E4 are spiny stellate cells; B in E5B,E2B=bursting, R in E5R=regular firing; L in I2L,I6L=LTS=low threshold spiking (bursting and LTS not currently explicitly implemented)

#### Dynamics

We're interested in which wiring predisposes to sustained activity (ictal activity). So far we have only explored the Martin model. Some wirings show sustained activity. Initial stimulus is to E4 and E6 (thalamocortical inputs)

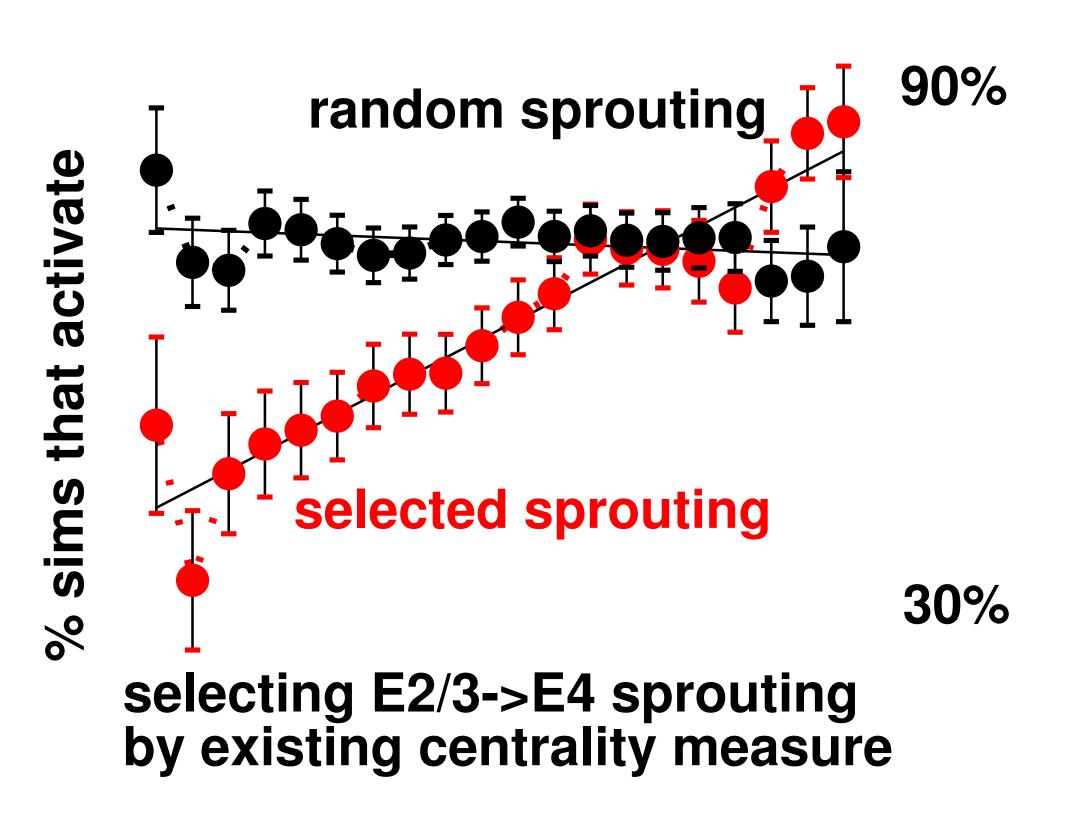


Cell model

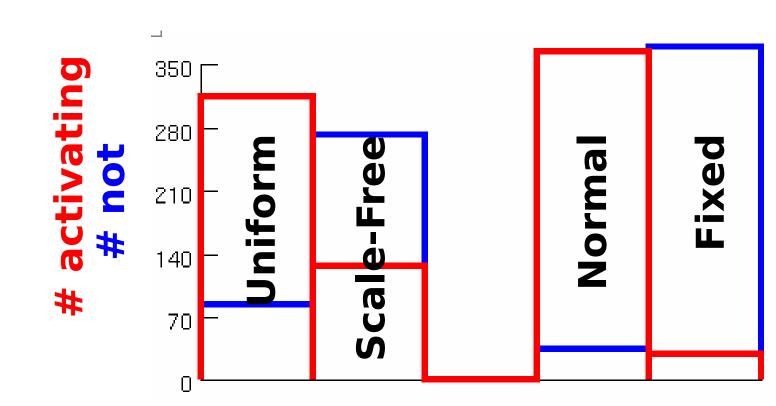


### Sprouting

We predicted that sprouting along pathways involving E2/3 would increase number of hubs and thereby promote network activation. Tested by either sprouting from randomly chosen cells (black) or from cells chosen for existing low centrality (red left) or existing high centrality (red right).



Haven't solved problem of balancing weights so as to get 50/50 active/not-active. Here we use different connectivity algorithms (within or between layers) but do not get near this balance (33/66 for scale-free case). We may need to use an adaptive algorithm to set weights. (See Poster Column 1 for description of the 4 distributions used.)



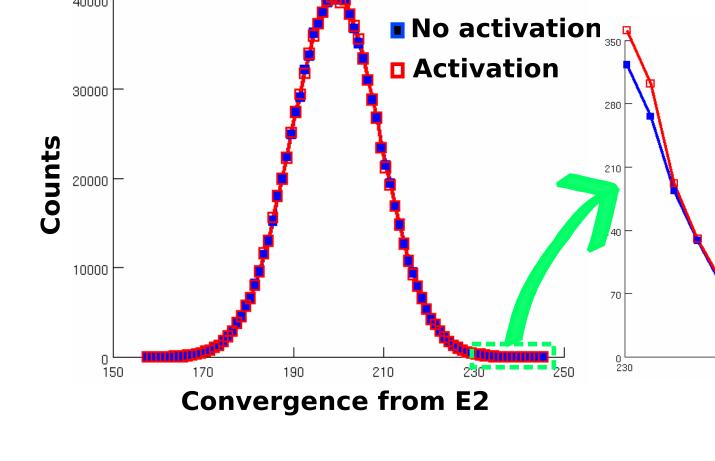
How do the activated networks (red) differ

marginal and subtle -- ie a small increase

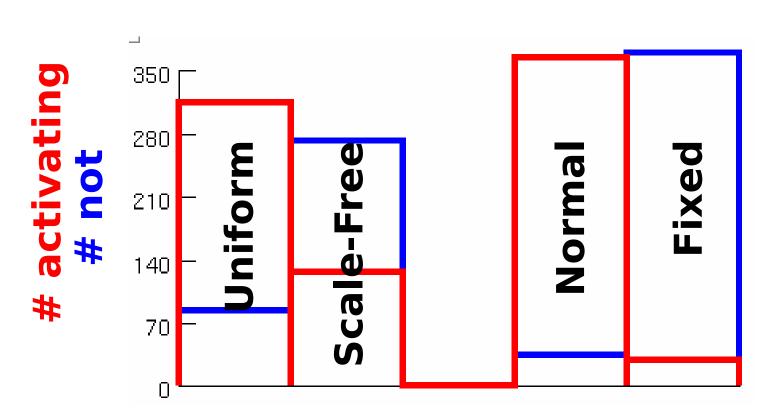
from the not-activated (blue). Differences are

in hubs may make a big difference. Here we look at

convergence from E2/3 cells for all units to which



To look at multiple factors, we ask how well activation/not predicts unusually high (red) or unusually low (blue) measures. Effect strength is given by circle diameter for different layers (x-axis) across different measures (y-axis). Internal E2/3 (E2-E2) convergence and multiple measures into E5 are [E58] important. (We excluded here the NORMAL group where almost all networks activated as shown in histogram above.)



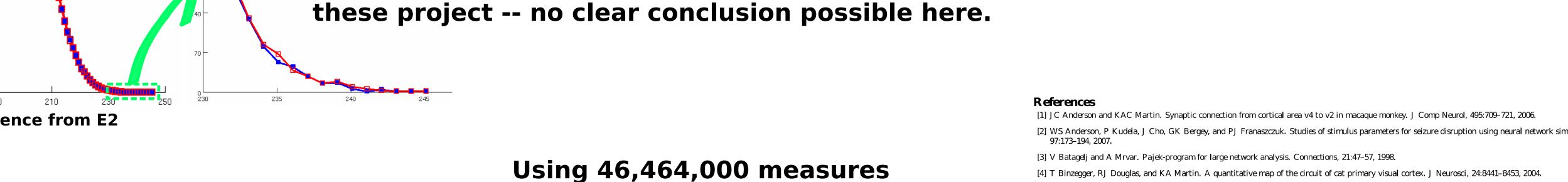
1600 simulations run

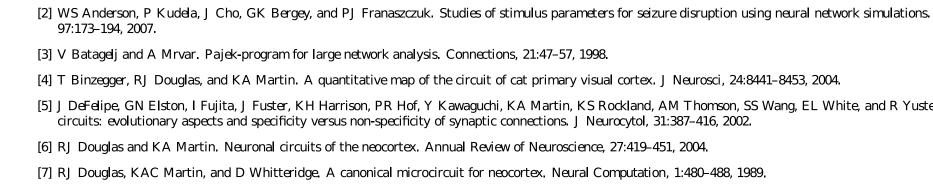
## 1. L2/3 excitatory cells provide a central

hub enabling cortical activation. 2. Reciprocal excitatory feedback between L5 and L2/3 also critical.

Conclusions

- 3. Standard neurobiological measures of type-specific convergence and divergence are more useful than general graph-theoretic measures in predicting dynamics.
- 4. Small variation in degree distribution in critical cell populations (layers) produces large changes in dynamics.





- [17] A Noack. An energy model for visual graph clustering. Lecture notes in computer science, pages 425-436, 2003
- [20] A Roxin, H Riecke, and SA Solla. Self-sustained activity in a small-world network of excitable neurons. Physical Review Letters, 92:198101, 2004. [22] O Sporns. Small-world connectivity, motif composition, and complexity of fractal neuronal connections. Biosystems, 85:55-64, 2006.
- [24] RD Traub, D Contreras, MO Cunningham, H Murray, FE LeBeau, A Roopun, A Bibbig, WB Wilent, MJ Higley, and MA Whittington. Single-column thalamocortical network model exhibiting gamma oscillations, sleep spindles, and epileptogenic bursts. J Neurophysiol, 93:2194-2232, 2005. [25] N Weiler, L Wood, J Yu, SA Solla, and GM Shepherd. Top-down laminar organization of the excitatory network in motor cortex. Nat Neurosci, 11:360-366, 2008.